# PLIER:Pathway-Level Information Extractor 

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## Generative model for gene expression data

- Gene expression is driven by upstream factors that give rise to the observed data structure.
- PCA gives us a representation of these upstream factors but not a one-to-one correspondence.


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$$
\text { MINIMIZE } \quad\|Y-Z B\|_{F}^{2}+\lambda\|Z\|_{L^{1}}
$$

SUBJECT TO $Z>0$.

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 recover some, but not all, variables of interest.
- These methods are data agnostic, they don't make use of gene identities!
- We want not just the most parsimonious but also the most biologically meaningful decomposition.


## PLIER: Pathway-Level Information ExtractoR

Idea: Make use of gene identities.

SUBJECT TO $\quad \operatorname{rank}(Z)=k, \quad \operatorname{rank}(B)=k, \quad U>0, \quad Z>0$.


Prior knowledge matrix $C$ is a binary geneset representation, where each column is a potentially co-regulated set of genes. Number of genesets is many times larger than $k$.

## Implementation Details

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- All constants are set automatically
- Running time depends on the size of the data and size of $C$
- We pre-compute the inverse of $C$ and use it to find a set of active genesets in each iteration to be optimized with the elastic-net penalty


## Recovering the pathway effects with PLIER

Revisit the toy example composition variables


Method
Ideal
PCA
Sparse positive

## Recovering the pathway effects with PLIER

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## Recovering the pathway effects with PLIER

- Performance across repeated simulations
Revisit the toy example composition variables

- Recovering 30 pathway effects from a prior information database of 1000 pathways



## How do we use PLIER?

Example on real human blood dataset ( 35 samples) with directly measured by Cytof


U matrix for a large dataset (DGN)

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```
SVM T cells regulatory (Tregs)
KEGG_SPLICEOSOME
```

KEGG CHRONIC MYELOID LEUKEMIA
MIPS TFTC_TYPE HISTONE ACETYL TRANSFERASE_COMPLEX
REACTOME RESPIRATORY ELECTRON_TRANSPORT
BIOCARTA CDC42RAC PATHWAY
KIOCARTA CDC
REACTOME RNA POL I PROMOTER_OPENING
BIOCARTA_ PROTEASOME_PATHWAY
MIPS 26S PROTEASOME
REACTOME_ACTIVATED_AMPK_STIMULATES_FATTY_ACID_OXID
KEGG DNA REPLICATION
MIPS_EIF3_COMPLEX
MIPS 40S RIBOSOMAL_SUBUNIT_CYTOPLASMIC
KEGG RIBOSOME
REACTOME_GENERIC_TRANSCRIPTION_PATHWAY REACTOME FORMATION OF ATP BY CHEMIOSMOTIC COUPLING REACTOME FORMATION OF ATP BY C KEGG-LYSOSOME
I TCELLA7
TCELLA6
TCELLA4
TCELLA2
TCELLA
MEGA2
MEGA2
MEGA1
DENDA1
NKcell-control
Monocyte-Day0
Bcell-Memory_IgM
Bcell-naïve

$$
-\quad \stackrel{\infty}{0} \stackrel{\circ}{0} \dot{0}
$$

## U matrix for a large dataset (DGN)



- How do we know the pathways are real? we zero-out a random $1 / 5$ of the genes for every pathway before optimization and check if we get them back in the loading.


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- We can see many cell types.
- 3 kinds of CD8 T cells.
- Naive and memory B-cells.
- Very high frequency cell-types have multiple LVs.


## How do we use PLIER?

PLIER latent variables can be plugged into any downstream analysis that would normally be done at the gene level-for example eQTLs.

| LV id | LV name | snps | cis-Gene(s) | corrected p-value |
| ---: | :--- | :--- | :--- | :--- |
| 44 | Mega/platelet 1 | rs1354034 | ARHGEF3 | $<1.45 \mathrm{e}-10$ |
| 133 | Mega/platelet 2 | rs1354034 | ARHGEF3 | 0.01547 |
| 120 | Histones | rs1354034 | ARHGEF3 | 0.01889 |
| 97 | Zinc fingers, pseudogenes | rs1471738 | SENP7 | $<1.45 \mathrm{e}-10$ |
| 56 | PLAGL1 associated, myeloid | rs9321957 | PLAGL1 | $3.6 \mathrm{e}-05$ |
| $42^{*}$ | IKZF1 associated, myeloid | rs10251980 | IKZF1 | $<1.45 \mathrm{e}-10$ |
| 17 | NEK6 associated, myeloid | rs16927294 | NEK6 | 0.00360 |
| 67 | Neutrophils | rs13289095 | PKN3,SET,ZDHHC12 | 0.01888 |
| $55^{*}$ | NFE2 associated, erythrocyte | rs35979828 | NFE2 | $<1.45 \mathrm{e}-10$ |
| 21 | Interferon-gamma | rs3184504 | SH2B3 | $5.9 \mathrm{e}-05$ |
| 40 | NFKB/TNF | rs12100841 | PPP2R3C | 0.00204 |
| 16 | Myeloid/ILC | rs1138358 | BCL2A1,MTHFS,ST20 | 0.00025 |

Interferon-gamma LV21 uses 3 pathways:

- REACTOME_INTERFERON_GAMMA_SIGNALING
- GSE19182 lfng
- SANA_RESPONSE_TO_IFNG_UP


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## A single locus controls 2 pathway effects

## LV eQTLs pathway associations

|  |  |  |  |  |  |  |  |  |  | WIERENGA_STAT5A_TARGETS_DN |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
|  |  |  |  |  |  |  |  |  |  | MEGA2 |
|  |  |  |  |  |  |  |  |  |  |  |
| RAGHAVACHARI_PLATELET_SPECIFIC_GENES |  |  |  |  |  |  |  |  |  |  |

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Platelet

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WIERENGA_STAT5A_TARGETS_DN
MEGA2
RAGHAVACHARI_PLATELET_SPECIFIC_GENES CREIGHTON_ENDOCRINE_THERAPY_RESISTANCE_2 SEKI_INFLAMMATORY_RESPONSE_LPS_UP LINDSTEDT_DENDRITIC_CELL_MATURATION_B GILMORE CORE NFKB_PATHWAY KEGG_SYSTEMIC_LUPUS_ERYTHEMATOSUS REACTOME_MEIOTIC_RECOMBINATION REACTOME_RNA_POL_I_PROMOTER_OPENING SANA_RESPONSE_TO_IFNG_UP REACTOME_INTERFERON_GAMMA_SIGNALING GSE19182_lfng REACTOME_GENERIC_TRANSCRIPTION_PATHWAY

Top genes for mega/platelet LVs \% $\dagger$ ion $+\infty$ Z-score ITGB5 SPARC CLU ITGA2B ITGB3 ALOX12

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ITGB5 SPARC

LV133 (Mega/platelet LV early) genes are expression in megakaryocyte precursors.
LV44 (Mega/platelet LV late) genes are megakaryocyte specific.

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WIERENGA_STAT5A_TARGETS_DN

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MEGA2
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RAGHAVACHARI_PLATELET_SPECIFIC_GENES
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SEKI_INFLAMMATORY_RESPONSE_LPS UP
LINDSTEDT_DENDRITIC_CELL_MATURATION_B GILMORE CORE NFKB_PATHWAY KEGG_SYSTEMIC_LUPUS_ERYTHEMATOSUS REACTOME_MEIOTIC_RECOMBINATION REACTOME_RNA_POL_I_PROMOTER_OPENING SANA_RESPONSE_TO_IFNG_UP REACTOME_INTERFERON_GAMMA_SIGNALING GSE19182_lfng REACTOME_GENERIC_TRANSCRIPTION_PATHWAY NAKAYAMA_SOFT_TISSUE_TUMORS_PCA1_UP NKA1

## PID_PRLSIGNALINGEVENTSPATHWAY

Neutrophil-Resting
MARTINELLI_IMMATURE_NEUTROPHIL_UP NKcell-control

厤 ERY2


Mega/platelet LV 133
Mega/platelet LV 44

Top genes for mega/platelet LVs
1 ○ $\mathrm{N} \rightarrow \mathrm{m}$ Z-Score
rs1354034--ARGHEF associations


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## Pleitropy of the ARGEF3 locus

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| phenotype | reported SNP | Close gene | LV 133 p-value | LV44 p-value | proxy SNP |
| ---: | :--- | :--- | :--- | :--- | :--- |
| PLT | rs2911132 | ERAP2 | $\mathbf{2 . 4 4 1 7 e - 0 5}$ | 0.13817361 | rs2549803 |
| MPV | rs10876550 | COPZ1 | 0.69933 | $\mathbf{1 . 1 8 4 7 e - 0 5}$ | rs10876550 |

Table: Raw p-values. 80 platelet related SNPs tested.

[^0]
## PLIER models transfer across datasets

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PLIER decompositions performed independently


NESDA LVs
DGN top pathway

## Correlation with phenotypes is more consistent in LV space



## Some fun results

- Dataset from a collaborator: melanoma RNAseq , immunotherapy reponse (8 progressors, 11 responders).
- Very similart to the published Hugo et al. dataset * (13 progressors, 15 responders). How do they compare?


[^1]
## Usoskin et al. dataset

## scRNAseq of mouse sensory neurons.



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- Minimally supervised method: selects relevant pathways and discards thousands of irrelevant ones.
- Additional output matrix $U$ provides the mapping between pathways and LVs for quick interpretation.
- Pathway-level estimates can be used in any subsequent analysis yielding mechanistic hypotheses.


## Questions and future directions

- Group-level regularization on samples: not every LV exists in every sample.


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- Looking for LVs that maximize objectives other than variance.
-When are positivity constraints on the loadings necessary?


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- U54 HG008540-03
- 1R03 MH109009-01A1


[^0]:    Furman-Niedziejko A. et al. Relationship between abdominal obesity, platelet blood count and mean platelet volume in patients with metabolic syndrome.

[^1]:    * Genomic and Transcriptomic Features of Response to Anti-PD-1 Therapy in Metastatic Melanoma

